



**CAMOSUN COLLEGE**  
**School of Arts & Science**  
**Department of Chemistry & Geoscience**

**CHEM-255-D01**  
**Biochemistry**  
**Winter Semester, 2021**

**COURSE OUTLINE**

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The course description is online @ <http://camosun.ca/learn/calendar/current/web/chem.html>

\* Please note: This outline will not be kept indefinitely. It is recommended students keep this outline for their records, especially to assist in transfer credit to post-secondary institutions.

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**1. Instructor Information**

<b>(a) Instructor</b>	Jamie Doran, Ph.D.
<b>(b) Office hours</b>	Tuesday 1:30 – 2:20 PM Wednesday 1:30 – 2:20 PM Thursday 1:30 – 2:20 PM Friday 1:30 – 3:20 PM
<b>(c) Location</b>	On-line. Arrange via email to meet via Zoom. Available at other times.
<b>(d) Phone</b>	250.370.3441 (email preferable since on-line)
<b>(e) E-mail</b>	<a href="mailto:jdoran@camosun.ca">jdoran@camosun.ca</a> (available evenings and weekends throughout the semester)

**2. Intended Learning Outcomes**

Upon completion of this course the student will be able to:

1. Describe the fundamental characteristics of proteins, including enzymes, carbohydrates, lipids, and the nucleic acids, DNA and RNA.
2. Describe the chemical and biochemical principles governing the structure-function relationships of biomolecules and membranes.
3. Describe the underlying themes of key biochemical processes, including metabolism, bioenergetics, gene expression and protein synthesis.
4. Describe and evaluate the important aspects of the biochemical framework of cellular function at a molecular level, and the chemical bases thereof.
5. Use the basic vocabulary of biochemistry.
6. Explain the significance of biochemistry in clinical and veterinary medicine, laboratory analyses, nutrition, agriculture, and biotechnology.
7. Isolate specific proteins using gel-permeation, ion-exchange, and affinity-based column chromatography methods.
8. Analyze proteins by SDS-polyacrylamide gel electrophoresis.
9. Conduct polymerase chain reaction amplification of DNA molecules.
10. Utilize agarose gel electrophoresis for analysis of DNA samples.
11. Critically analyze the results obtained using each of the biochemical experimental techniques described above.
12. Work with an awareness of the basic safety considerations and general procedures associated with a biochemistry laboratory.

### 3. Required Materials

#### (a) Texts

**Moran, L.A., Horton, H.R., Scrimgeour, K.G. & Perry, M.D. 2012. *Principles of Biochemistry*. 5<sup>th</sup> ed. Pearson Education Inc., Toronto.**

This textbook is *absolutely required* for this course. It may be purchased from the Lansdowne Campus bookstore. Chapter study guides are included in the course pack also containing the laboratory manual (please see below). Links to relevant web-based learning resources are presented in the text. A collection of relevant lecture slides, largely based on this textbook, are available as a separate course pack (please see below).

#### (b) Other

##### **Chem 255 Laboratory Manual & Course Study Guides.**

This course pack contains experimental procedures with introductory material that, along with the textbook, provide an understanding of the biochemical techniques employed in the laboratory component of the course. This course pack also includes chapter study guides for optimal use of the textbook and lecture notes. *This course pack is required material* and is available through the Lansdowne Campus bookstore. It is also posted at the course website on D2L.

##### **Chem 255 Lecture Slides Course Package.**

This *required material* has proven to be vital in promoting optimal lecture-based learning, in-class discussion, and salient notetaking. It works optimally as a hardcopy. It is available through the Lansdowne Campus bookstore. It is also posted at the course website on D2L.

#### **General Materials and Supplies**

Computer Each student is required to have a computer or other device for use in on-line classes, office hours and exams. It is highly recommended that this device be a computer, or a tablet fitted with a keyboard. A mobile phone may be used but it is anticipated that students will be able to work very effectively on-line when participating in classes, term tests and the final exam.

Calculator A basic scientific calculator is *required* at times for work involving experimental procedures and lecture material, and may be required during term tests and the final exam. Each student is *required* to provide her or his own calculator. Cell phone-based, tablet-based, or computer-based calculators are sufficient during this on-line offering of the course. Graphing calculators are not to be used during term tests or the final exam.

### 4. Course Content and Schedule

Credits	4 credits
In-class workload	6 hours per week in a synchronous offering scheduled by the College  There are four 50-min lectures per week (please see below). Term test review periods will be scheduled into appropriate lecture periods prior to each term test.  Laboratory periods will be used to discuss experiments and experimental techniques in biochemistry. These scheduled periods are also used to host two term tests (please see below).
Out-of-class workload	6 hours per week
Number of weeks	14 weeks
Pre-requisite	C in Chem 121 - College Chemistry 2
Pre- or Co-requisite	Chem 230 – Organic Chemistry 1

#### **Course times and locations**

<u>Lectures</u>	Monday, 8:30 AM - 9:20 AM Wednesday, 8:30 AM - 9:20 AM Thursday, 8:30 AM - 9:20 AM Friday, 8:30 AM - 9:20 AM
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Laboratory Periods & Term Test Times Please refer to the laboratory and term test schedule, below.

Tuesday, 8:30 AM to 10:20 AM on-line

## **Lecture Outline**

A general outline of the topics to be covered in the course is provided below. Study guides for each chapter of the textbook are provided in the *Chemistry 255 - Biochemistry Laboratory Manual and Course Study Guides* course pack.

### **Introduction to Biochemistry**

[Chapter 1]

Introduction; Perspective on early developments in biochemistry; Biochemically Relevant Elements; Classes of Organic Compounds, Functional Groups & Linkages; SI unit prefixes and metric conversions

### **Noncovalent bonding, pH & pKa, and Buffers & Buffering**

[Chapter 2]

Noncovalent Bonding in Biomolecules; Buffers & Buffering; pH & pKa; The Henderson-Hasselbach Equation; The Bicarbonate Blood-Buffer System.

### **Amino Acids and the Primary Sequence of a Protein**

[Chapter 3]

Structures of Common Amino Acids; Ionization and Potential Roles of Amino Acid Functional Groups in Covalent and Noncovalent bonding; Peptide Bonds; Peptide Bond Groups and Related Structure-Function Relationships; Protein Purification Techniques; Protein Sequencing; MALDI-TOF MS; Comparative Analyses of Protein Sequences.

### **Protein Structure & Function**

[Chapter 4]

The Nature of the Peptide Bond; Secondary, Super-Secondary, Tertiary and Quaternary Protein Structures; Protein Folding, Denaturation and Renaturation; Protein Structure-Function Relationships.

### **Enzymes**

[Chapter 5]

Classes of Enzymes; Enzyme Kinetics; Enzyme Inhibition; Lineweaver-Burk Plots; Allosteric and Covalent Regulation of Enzyme Activity.

### **Mechanisms of Enzyme Catalysis**

[Chapter 6]

Overview of Enzyme Function; Chemical Mechanisms of Enzyme Catalysis; Proximity Effects; Transition State Stabilization; Mechanism of Chymotrypsin Catalysis.

### **Coenzymes and Vitamins**

[Chapter 7]

Essential Ions, Vitamins and Coenzymes - Structures and Roles

### **Carbohydrates**

[Chapter 8]

Structures and Functions of Monosaccharides, Disaccharides, Polysaccharides, Proteoglycan Complexes and Glycoconjugates.

### **Lipids & Membranes**

[Chapter 9]

Classes of Lipids; Structures and Roles of Lipids; Membrane Structure/Function; Membrane Transport; Transmembrane Signal Transduction.

### **Overview of Metabolism**

[Chapter 10]

Introduction to Intermediary Metabolism and Bioenergetics.

### **Glycolysis**

[Chapter 11]

The Nature and Role of Glycolytic Metabolic Pathway, and its Regulation.

### **Other Major Pathways in Carbohydrate Metabolism**

[Chapter 12]

Gluconeogenesis; Cori Cycle; Glucose-Alanine Cycle; Pentose Phosphate Pathway; Glycogen Metabolism; Maintenance and Regulation of Blood Glucose Levels.

### **The Citric Acid Cycle**

[Chapter 13]

Pyruvate Dehydrogenase (PDH) Complex Activity; The Nature and Roles of Krebs Cycle (aka TCA Cycle, Citric Acid Cycle); Regulation of the PDH Complex.

### **Electron Transport and Oxidative Phosphorylation**

[Chapter 14]

The Chemiosmotic Hypothesis; Electron Transport; Oxidative Phosphorylation; Shuttling of Krebs Cycle Intermediates; Glycerol Phosphate & Malate-Aspartate Shuttle Systems.

### **Lipid Metabolism**

[Chapter 16]

Lipoprotein Structure and Function; Mobilization of Fatty Acids;  $\beta$ -Oxidation; Cholesterol and Ketone Metabolism.

### **Amino Acid Metabolism**

[Chapter 17]

Assimilation of Ammonia; Amino Acid Catabolism and Anabolism; Renal Glutamine Metabolism.

### **Nucleotide Metabolism**

[Chapter 18]

Purine and Pyrimidine Biosynthetic Pathways.

Template Published by Educational Approvals Office (VP Ed Office)

1/1/2021

## **DNA & RNA**

[Chapter 19]

Nucleosides & Nucleotides; DNA Structure; Restriction Endonucleases and DNA Cloning; Physical Mapping of DNA; DNA Fingerprinting.

## **DNA Replication and Biotechnological Applications**

[Chapter 20]

DNA Sequencing; Site-Specific Mutagenesis; Polymerase Chain Reaction (PCR).

## **RNA Synthesis (Transcription)**

[Chapter 21]

RNA Polymerase Function; Promoters; Transcriptional Regulation of the *lac* Operon.

## **Protein Synthesis (Translation)**

[Chapter 22]

The Shine-Dalgarno Sequence & the Initiation of Translation; Translational Regulation by Repression & Attenuation; Signal Sequences and Protein Secretion.

## **Laboratory & Term Test Schedule**

*In preparation, please read through the introductory material in the laboratory manual & refer to relevant material in the textbook. Preparation of a flow chart for each experiment is required. Please complete these and, when appropriate, the pre-lab questions in time for submission at each laboratory period.*

Tuesday, January 12<sup>th</sup> **Introduction to D2L, for anyone unfamiliar with it.**

Tuesday, January 19<sup>th</sup> **How to make a Tris-HCl buffer**

Tuesday, January 26<sup>th</sup> **Experiment 1  
Introduction to Protein Purification by Column Chromatography  
Gel Permeation Column Chromatography**

Tuesday, February 2<sup>nd</sup> **Experiments 2 & 3  
Purification of Proteins by Ion-Exchange Column Chromatography  
& by Affinity Column Chromatography**

Tuesday, February 9<sup>th</sup> **Experiment 4  
SDS-Polyacrylamide Gel Electrophoresis (SDS-PAGE) Part 1.**

Tuesday, February 16<sup>th</sup> **Reading Break**

Tuesday, February 23<sup>rd</sup> **Term Test 1**

Tuesday, March 2<sup>nd</sup> **Experiment 4 - SDS-PAGE Part 2.**

Tuesday, March 9<sup>th</sup> **Experiment 5 Polymerase Chain Reaction (PCR)**

Tuesday, March 16<sup>th</sup> **Site-Directed Mutagenesis (SDM); Gene Cloning**

Tuesday, March 23<sup>rd</sup> **Sanger Enzymatic DNA Sequencing**

Tuesday, March 30<sup>th</sup> **Term Test 2**

Tuesday, April 6<sup>th</sup> **RFLP-based DNA testing.**

Tuesday, April 13<sup>th</sup> **Final Exam Review**

**Final Exam** The date and time will be published by the College during the semester.

## **5. Basis of Student Assessment (Weighting)**

### (a) Assignments

#### **Metabolic Pathways Chart Project.**

This assignment is described in detail in a handout to be provided once topics of intermediary metabolism arise in the course. Everyone is required to hand in the results of her or his own work. This 'Metabolic Pathways' chart is due on the final day of class. This project contributes **5%** to the final grade.

(b) Term Tests

**Term Test #1**

This test covers relevant material from approximately the first third of the course. The delineation of material that students are responsible for, including that from the laboratory section of the course, will be provided in class about one week before the date of the test. This is a 110-minute test that will be written on Tuesday, February 23<sup>rd</sup> from 8:30 AM to 10:20 AM. The results of this test contribute **20%** of the final grade.

**Term Test #2**

This test covers relevant material from approximately the second third of the course. The delineation of material that students are responsible for, including that from the laboratory section of the course, will be provided in class about one week before the date of the test. This is a 110 min. test that will be written on Tuesday, March 30<sup>th</sup> from 8:30 AM to 10:20 AM. The results of this test contribute **20%** of the final grade.

*If either term test is missed due to illness, or a similarly justifiable reason, with accompanying documentation the percentage value of that term test will be added to the value of the final exam.*

(c) Final Exam

The final exam is a comprehensive exam that includes components from the laboratory section of the course. The value this exam contributes to the final grade is **35%**. The time and location of the final exam will be published by the College during the semester.

**Attendance at the final exam is mandatory.** Appropriate documentation must accompany any explanation for absence if an I-grade (incomplete) is warranted.

(d) Other

*On-line laboratory period attendance is mandatory.* Pre-lab or post-lab assignments contribute equally a total of **15%** of the final grade.

Understanding of the principles, scientific and technical bases of each experiment is subject to examination on term tests and the final exam.

On-line lecture attendance and participation contributes **5%** to the final grade.

## 6. Grading System

Standard Grading System (GPA)

Competency Based Grading System

## 7. Recommended Materials to Assist Students to Succeed Throughout the Course

Please refer to descriptions of the required textbook, and required course packages, in Section 3, above.

Within D2L are tutorials that provide instruction on the use of Brightspace.

## 8. College Supports, Services and Policies



### Immediate, Urgent, or Emergency Support

If you or someone you know requires immediate, urgent, or emergency support (e.g. illness, injury, thoughts of suicide, sexual assault, etc.), **SEEK HELP**. Resource contacts

@ <http://camosun.ca/about/mental-health/emergency.html> or <http://camosun.ca/services/sexual-violence/get-support.html#urgent>

### College Services

Camosun offers a variety of health and academic support services, including counselling, dental, disability resource centre, help centre, learning skills, sexual violence support & education, library, and writing centre. For more information on each of these services, visit the **STUDENT**

**SERVICES** link on the College website at <http://camosun.ca/>

### College Policies

Camosun strives to provide clear, transparent, and easily accessible policies that exemplify the college's commitment to life-changing learning. It is the student's responsibility to become familiar with the content of College policies. Policies are available on the College website at <http://camosun.ca/about/policies/>. Education and academic policies include, but are not limited to, Academic Progress, Admission, Course Withdrawals, Standards for Awarding Credentials, Involuntary Health and Safety Leave of Absence, Prior Learning Assessment, Medical/Compassionate Withdrawal, Sexual Violence, Student Ancillary Fees, Academic Integrity, Grade Review & Appeals, Student Misconduct and Academic Accommodations for Students with Disabilities and Student Penalties and Fines.

## A. GRADING SYSTEMS <http://camosun.ca/about/policies/index.html>

The following two grading systems are used at Camosun College:

### 1. Standard Grading System (GPA)

Percentage	Grade	Description	Grade Point Equivalency
90-100	A+		9
85-89	A		8
80-84	A-		7
77-79	B+		6
73-76	B		5
70-72	B-		4
65-69	C+		3
60-64	C		2
50-59	D		1
0-49	F	Minimum level has not been achieved.	0

## 2. Competency Based Grading System (Non-GPA)

This grading system is based on satisfactory acquisition of defined skills or successful completion of the course learning outcomes

Grade	Description
COM	The student has met the goals, criteria, or competencies established for this course, practicum, or field placement.
DST	The student has met and exceeded, above and beyond expectation, the goals, criteria, or competencies established for this course, practicum, or field placement.
NC	The student has not met the goals, criteria or competencies established for this course, practicum, or field placement.

## B. Temporary Grades

Temporary grades are assigned for specific circumstances and will convert to a final grade according to the grading scheme being used in the course. See Grading Policy at <http://camosun.ca/about/policies/index.html> for information on conversion to final grades, and for additional information on student record and transcript notations.

Temporary Grade	Description
I	<i>Incomplete:</i> A temporary grade assigned when the requirements of a course have not yet been completed due to hardship or extenuating circumstances, such as illness or death in the family.
IP	<i>In progress:</i> A temporary grade assigned for courses that are designed to have an anticipated enrollment that extends beyond one term. No more than two IP grades will be assigned for the same course.
CW	<i>Compulsory Withdrawal:</i> A temporary grade assigned by a Dean when an instructor, after documenting the prescriptive strategies applied and consulting with peers, deems that a student is unsafe to self or others and must be removed from the lab, practicum, worksite, or field placement.

### Academic Honesty

Please become familiar with the School of Arts & Science guide on academic honesty:

<http://camosun.ca/learn/school/arts-science/images/Arts%20and%20Science%20Academic%20Honesty%20Guidelines.pdf>

Based on College policy, the outcome of an incident of academic honesty on an evaluation (lab-related submission, term test, exam) is a **failing grade** for that evaluation.

### Please Note:

Students may **not** use recording devices without the prior permission of the instructor or the Centre for Accessible Learning. The instructor's permission is not required when the use of a recording device is sanctioned by the College's Centre for Accessible Learning in order to accommodate a student's disability, *and* when the instructor has been provided with an instructor notification letter which specifies the use of a recording device. Such recordings are for the student's personal use only, and distribution of recorded material is prohibited. Recordings made during the course would include statements, questions and comments made by students in the class, and these are not to be disseminated or repeated in any manner based on the recordings. Otherwise, please have all recording devices turned off and put away while in lectures.