

CAMOSUN COLLEGE School of Arts & Science Department of Chemistry & Geoscience

CHEM-255-001 Biochemistry Winter 2019

COURSE OUTLINE

The course description is online @ http://camosun.ca/learn/calendar/current/web/chem.html

 Ω Please note: This outline will <u>not</u> be kept indefinitely. It is recommended students keep this outline for their records, especially to assist in transfer credit to post-secondary institutions.

1. Instructor Information

(a) Instructor	Jamie Doran, Ph.D.
(b) Office hours	Monday, 2:00 PM to 3:00 PM
	Tuesday, 2:00 PM to 3:00 PM
	Wednesday, 2:00 PM to 3:00 PM
	Thursday, 2:00 PM to 3:00 PM
	Friday, 2:00 PM to 3:00 PM
	Please make an appointment to ensure the time is available for you when you like.
	Students are welcome to request an appointment at other times.
(c) Location	Room 350C, Fisher Building, Lansdowne Campus
(d) Phone 250.3	370.3441 (voice-mail available)
(e) E-mail	jdoran@camosun.ca (available evenings and weekends throughout the semester)

2. Intended Learning Outcomes

Upon completion of this course the student will be able to:

- 1. Describe the fundamental characteristics of proteins, including enzymes, carbohydrates, lipids, and the nucleic acids, DNA and RNA.
- 2. Describe the chemical and biochemical principles governing the structure-function relationships of biomolecules and membranes.
- 3. Describe the underlying themes of key biochemical processes, including metabolism, bioenergetics, gene expression and protein synthesis.
- 4. Describe and evaluate the important aspects of the biochemical framework of cellular function at a molecular level, and the chemical bases thereof.
- 5. Use the basic vocabulary of biochemistry.
- 6. Explain the significance of biochemistry in clinical and veterinary medicine, laboratory analyses, nutrition, agriculture, and biotechnology.
- 7. Isolate specific proteins using gel-permeation, ion-exchange, and affinity-based column chromatography methods.
- 8. Analyze proteins by SDS-polyacrylamide electrophoresis.
- 9. Conduct polymerase chain reaction amplification of DNA molecules.
- 10. Utilize agarose gel electrophoresis for analysis of DNA samples.
- 11. Critically analyze the results obtained using each of the biochemical experimental techniques described above.
- 12. Work with an awareness of the basic safety considerations and general procedures associated with a biochemistry laboratory.

3. Required Materials

(a) Text

Moran, L.A., Horton, H.R., Scrimgeour, K.G. & Perry, M.D. 2012, *Principles of Biochemistry.* 5th ed. Pearson Education Inc., Toronto.

This textbook is *required* for this course. It may be purchased from the Lansdowne campus bookstore. Also, a copy is available in the Reserve Library at the Lansdowne campus. Chapter study guides are included in the course pack containing the laboratory manual (please see below). Links to relevant web-based learning resources are presented in the text. A collection of relevant lecture slides, primarily based on the textbook, are available as a separate course pack (please see below).

(b) Other

Chem 255 Laboratory Manual & Course Study Guides.

This course pack contains experimental procedures with introductory material that, along with the textbook, provide an understanding of the biochemical techniques employed in the laboratory component of the course. This course pack also includes chapter study guides for optimizing the use of the textbook and lecture notes. *This course pack is required material*, and is available through the Lansdowne campus book store.

Chem 255 Lecture Slides Course Package.

This *required material* has proven to be vital in promoting optimal lecture-based learning, in-class discussion, and salient note-taking. It is available through the Lansdowne campus book store.

General Materials and Supplies

- <u>Safety glasses</u> Safety glasses are required when handling hazardous chemicals, and are recommended when handling laboratory glassware. <u>Each student is required to provide her or his own pair of safety glasses</u>. Students lacking safety glasses when they are required will not be permitted in the laboratory.
- Lab coats are *required* for all experimental work in the laboratory. Each student is required to provide her or his own lab coat.
- Latex gloves Latex or alternate 'non-allergenic' gloves *will be available in the laboratory* and are to be used when appropriate to protect the skin from potentially hazardous chemicals or, much more often, to protect biochemicals from degradative enzymes found on the skin.
- <u>Calculator</u> A scientific calculator is *required* at times in the laboratory, in lecture, and during term tests and the final exam. Each student is *required* to provide her or his own scientific calculator. Cell phonebased, tablet-based or computer-based calculators cannot be used during term tests or the final exam.

4. Course Content and Schedule

	Credits In-class workload	4 credits 6 hours per week There are four 50-min lectures Term test review periods will be scheduled into an appropria slot prior to each term test. Experiments, pre-lab talks & post-lab analyses are conducted most of the 1 h & 50 min laboratory periods. This time period used to host two term tests.	ate lecture during
	Out-of-class workload Number of weeks Pre-requisite Pre- or Co-requisite	6 hours per week 14 weeks C in Chem 121 - College Chemistry 2 Chem 230 – Organic Chemistry 1	
Course	times and locations Lectures	Tuesday, 12:30 - 1:20 PM Fisher Building, Room F360	
		Wednesday, 12:30 - 1:20 PM Fisher Building, Room F360	
		Thursday, 12:30 - 1:20 PM Fisher Building, Room F360	
		Friday, 12:30 - 1:20 PM Fisher Building, Room F360	
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Lecture Outline

A general outline of the topics to be covered in the course is provided below. Study guides for each chapter of the textbook are provided in the Chemistry 255 - Biochemistry Laboratory Manual and Course Study Guides course pack. Each chapter study guide includes a section by section reading list inclusive of a listing of salient figures and tables, a listing of the relevant vocabulary, and recommended practice questions.

Introduction to Biochemistry

Introduction; Perspective on early developments in biochemistry; Biochemically Relevant Elements; Classes of Organic Compounds; Functional Groups; SI unit prefixes and metric conversions

Noncovalent bonding, pH & pKa, and Buffers & Buffering

Noncovalent Bonding in Biomolecules; pH & pKa; The Henderson-Hasselbach Equation; Buffers & Buffering: The Bicarbonate Blood-Buffer System.

Amino Acids and the Primary Sequence of a Protein

Structures of Common Amino Acids: pKa: Ionization and Potential Roles of Amino Acid Functional Groups in Covalent and Noncovalent bonding; Peptide Bonds; Peptide Bond Groups and Related Structure-Function Relationships; Protein Purification Techniques; Primary Protein Sequence; Protein Sequencing; MALDI-TOF MS; Comparative Analyses of Protein Sequences.

Protein Structure & Function

The Nature of the Peptide Bond; Secondary, Super-Secondary, Tertiary and Quaternary Protein Structures; Protein Folding, Stability, Denaturation and Renaturation; Protein Structure-Function Relationships.

Enzymes

Classes of Enzymes; Enzyme Kinetics; Enzyme Inhibition; Interpretation of Lineweaver-Burk Plots; Allosteric and Covalent Regulation of Enzyme Activity; Regulation of Quaternary-Structured Enzyme.

Mechanisms of Enzyme Catalysis

Overview of Enzyme Function: Chemical Mechanisms of Enzyme Catalysis; Proximity Effects and Transition State Stabilization; Mechanism of Chymotrypsin Activity.

Coenzymes and Vitamins

Roles and Structures of Essential Ions, Coenzymes, and Vitamins.

Carbohydrates

Roles and Structures of Monosaccharides, Disaccharides, Polysaccharides & Proteoglycan Complexes; Structure-Function Relationships in Carbohydrates and in Glycoconjugates.

Lipids & Membranes

Classes of Lipids, Structures and Roles of Lipids; Membrane Structure/Function; Membrane Transport; Transmembrane Signal Transduction Mechanisms.

Overview of Metabolism

Introduction to Intermediary Metabolism and Bioenergetics.

Glycolysis

The Nature and Role of Glycolytic Metabolic Pathway, and its Regulation.

Other Major Pathways in Carbohydrate Metabolism

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Gluconeogenesis; Cori Cycle; Glucose-Alanine Cycle; Pentose Phosphate Pathway; Glycogen Metabolism; Maintenance and Regulation of Blood Glucose Levels.

The Citric Acid Cycle

Dehydrogenase Complex Activity; The Nature and Roles of Citric Acid Cycle (aka TCA Cycle, Krebs Cycle); Regulation of the PDH Complex.

Electron Transport and Oxidative Phosphorylation [Chapter 14] Introduction to Bioenergetics; The Chemiosmotic Hypothesis; Electron Transport; Oxidative Phosphorylation in Mitochondria; Glycerol Phosphate & Malate-Aspartate Shuttle Systems; Shuttling of Krebs Cycle Intermediates.

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[Chapter 1]

[Chapter 2]

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[Chapter 4]

[Chapter 5]

[Chapter 6]

[Chapter 7]

[Chapter 8]

[Chapter 9]

[Chapter 11]

[Chapter 12]

[Chapter 13] Pyruvate

Phospholipid, Cholesterol and Ketone Bodies Metabolism; β -Oxidation.	-, , ,
Amino Acid Metabolism Assimilation of Ammonia; Amino Acid Catabolism and Anabolism; Renal Glutamine Metab	<i>[Chapter 17]</i> polism.
Nucleotide Metabolism Purine and Pyrimidine Biosynthetic Pathways.	[Chapter 18]
DNA & RNA Nucleosides & Nucleotides; DNA Structure; Restriction Endonucleases and DNA Cloning; DNA Finger-Printing.	[Chapter 19] Physical Mapping of DNA;
DNA Replication and Biotechnological Applications DNA synthesis; DNA Sequencing; Site-Specific Mutagenesis; Polymerase Chain Reactior	[Chapter 20] n (PCR).
RNA Synthesis (Transcription) RNA Polymerase Function; Promoters; Transcriptional Regulation of the <i>lac</i> Operon.	[Chapter 21]
Protein Synthesis (Translation) Ribosome Structure and Function; The Shine-Dalgarno Sequence & the Initiation of Trans	<i>[Chapter</i> 22 <i>]</i> slation; Translational

Lipoprotein Structure and Function; Storage and Mobilization of Fatty Acids and Cholesterol; Fatty Acid,

Laboratory & Term Test Schedule

Lipid Metabolism

In preparation, please refer to Chapter 3 in the text & the introductory material to each experiment. Preparation of a flow chart for each experiment is required. Please complete the full pre-lab assignments in time for submission at each laboratory period.

Thursday, January 10th Laboratory Orientation

Thursday, January 17th Mini-Experiment - Preparing a Tris Buffer

<u>Thursday, January 24th</u> Experiment 1 Separation of Proteins by Gel Permeation Column Chromatography

Regulation by Repression & Attenuation; Signal Sequences and Protein Secretion.

<u>Thursday, January 31st</u> Experiments 2 & 3 *Purification of Proteins by Ion-Exchange Column Chromatography* & Affinity Column Chromatography (start)

<u>Thursday, February 7th</u> Experiment 3 Isolation of Concanavalin A by Affinity Chromatography (completion) Discussion of Results, & Comparative Review of Chromatography Principles & Techniques

Thursday, February 14th Term Test 1 3:30 PM to 5:20 PM in F360

<u>Thursday, February 21st</u> Reading Week

<u>Thursday, February 28th</u> Experiment 4 **SDS-Polyacrylamide Gel Electrophoresis (SDS-PAGE): Separation & Identification of Proteins, and Determination of Protein Molecular Weight** Part 1 – Theory & Preparation of a Polyacrylamide Gel for the Separation of Proteins

Thursday, March 7th Experiment 4 - SDS-PAGE (continued) Part 2 - Protein Electrophoresis, & Staining for Detection of Separated Proteins

<u>Thursday, March 14th</u> **Experiment 4 - SDS-PAGE (completion)** Part 3 – Analyses of SDS-PAGE Results & Further Principles and Theory

Thursday, March 21st Term Test 2 3:30 PM to 5:20 PM in F360

<u>Thursday, March 28th</u> Experiment 5 **Polymerase Chain Reaction (PCR) Amplification of Cloned SAGE Tag Fragment, Part** 1 – Theory & PCR Amplification of DNA Fragments

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[Chapter 16]

<u>Thursday, April 4th</u> Experiment 5 (continuation) **PCR Amplification of Cloned SAGE Tag Fragments** Part 2 - Agarose Gel Electrophoresis & Detection of PCR Amplified DNA Fragments

 Thursday, April 11th
 Experiment 5 (completion)

 PCR Amplification of Cloned SAGE Tag Fragments

 Part 3 - Analysis of PCR Results

 \approx & Final Exam Review ~

s Final Exam a The time and location will be published by the College during the semester.

5. Basis of Student Assessment (Weighting)

(a) Assignments

Metabolic Pathways Chart Project.

This assignment is described in detail in a handout to be provided once topics of intermediary metabolism arise in the course. Each individual is required to hand in the results of her or his own work. This metabolic pathways chart is due on the final day of class. This project contributes **5%** to the final grade.

(b) Term Tests

Term Test #1

This test covers relevant material from approximately the first third of the course. The delineation of material that students are responsible for, including that from the laboratory section of the course, will be provided in class about one week before the date of the test. This is a 110-minute test that will be written on <u>Thursday</u>, <u>February 14th</u> from 3:30 PM to 5:20 PM in F360. The results of this test contribute to **20%** of the final grade.

Term Test #2

This test covers relevant material from approximately the second third of the course. The delineation of material that students are responsible for, including that from the laboratory section of the course, will be provided in class about one week before the date of the test. This is a 110 min. test that will be written on <u>Thursday, March 21st</u> from 3:30 PM to 5:20 PM in F360. The results of this test contribute to **25%** of the final grade.

If either term test is missed due to illness or similarly justifiable reason, with accompanying documentation the percentage value of that term test will be added to the value of the final exam.

(c) Final Exams

The final exam is a comprehensive exam that includes components from the laboratory section of the course. The value this exam contributes to the final grade is **40%**. The time and location of the final exam will be published by the College during the Fall Semester.

Attendance at the final exam is mandatory. Appropriate documentation must accompany any explanation for absence if an incomplete grade (I grade) is warranted.

(d) Other

Laboratory Experiments

Laboratory participation and performance contributes **5%** to the final grade. **Attendance in the lab periods is mandatory.** No laboratory experiment can be missed without an acceptable reason submitted in writing, such as a letter from a MD.

Pre-lab assignments and flowcharts also contribute equally a total of 5% of the final grade.

Please come to each lab period prepared for the experiment. Prepare a flow-chart and have the rest of the pre-lab assignment completed. Understanding of the principles, scientific and technical bases, and results of each experiment is subject to examination on term tests and the final exam.

6. Grading System

Standard Grading System (GPA)



Competency Based Grading System

7. Recommended Materials to Assist Students to Succeed Throughout the Course

Please refer to the required textbook and required course packages described above.

8. College Supports, Services and Policies



Immediate, Urgent, or Emergency Support

If you or someone you know requires immediate, urgent, or emergency support (e.g. illness, injury, thoughts of suicide, sexual assault, etc.), **SEEK HELP**. Resource contacts @ http://camosun.ca/about/mental-health/emergency.html or http://camosun.ca/services/sexual-violence/get-support.html#urgent

College Services

Camosun offers a variety of health and academic support services, including counselling, dental, disability resource centre, help centre, learning skills, sexual violence support & education, library, and writing centre. For more information on each of these services, visit the **STUDENT SERVICES** link on the College website at http://camosun.ca/

College Policies

Camosun strives to provide clear, transparent, and easily accessible policies that exemplify the college's commitment to life-changing learning. It is the student's responsibility to become familiar with the content of College policies. Policies are available on the College website at http://camosun.ca/about/policies/. Education and academic policies include, but are not limited to, Academic Progress, Admission, Course Withdrawals, Standards for Awarding Credentials, Involuntary Health and Safety Leave of Absence, Prior Learning Assessment, Medical/Compassionate Withdrawal, Sexual Violence and Misconduct, Student Ancillary Fees, Student Appeals, Student Conduct, and Student Penalties and Fines.

A. GRADING SYSTEMS http://camosun.ca/about/policies/index.html

The following two grading systems are used at Camosun College:

1. Standard Grading System (GPA)

Percentage	Grade	Description	Grade Point
			Equivalency
90-100	A+		9
85-89	А		8
80-84	A-		7
77-79	B+		6
73-76	В		5
70-72	B-		4
65-69	C+		3
60-64	С		2
50-59	D		1
0-49	F	Minimum level has not been achieved.	0

2. Competency Based Grading System (Non GPA)

This grading system is based on satisfactory acquisition of defined skills or successful completion of the course learning outcomes

Grade	Description
СОМ	The student has met the goals, criteria, or competencies established for this course, practicum or field placement.
DST	The student has met and exceeded, above and beyond expectation, the goals, criteria, or competencies established for this course, practicum or field placement.
NC	The student has not met the goals, criteria or competencies established for this course, practicum or field placement.

B. Temporary Grades

Temporary grades are assigned for specific circumstances and will convert to a final grade according to the grading scheme being used in the course. See Grading Policy at http://camosun.ca/about/policies/index.html for information on conversion to final grades, and for additional information on student record and transcript notations.

Temporary	Description
Grade	
I	<i>Incomplete</i> : A temporary grade assigned when the requirements of a course have not yet been completed due to hardship or extenuating circumstances, such as illness or death in the family.
IP	<i>In progress</i> : A temporary grade assigned for courses that are designed to have an anticipated enrollment that extends beyond one term. No more than two IP grades will be assigned for the same course.
CW	<i>Compulsory Withdrawal</i> : A temporary grade assigned by a Dean when an instructor, after documenting the prescriptive strategies applied and consulting with peers, deems that a student is unsafe to self or others and must be removed from the lab, practicum, worksite, or field placement.

Please Note:

Students may **not** use recording devices in the classroom without the prior permission of the instructor or The Centre for Accessible Learning. The instructor's permission is not required when the use of a recording device is sanctioned by the College's Centre for Accessible Learning in order to accommodate a student's disability, *and* when the instructor has been provided with an instructor notification letter which specifies the use of a recording device. Such recordings made in the classroom are for the student's personal use only, and distribution of recorded material is prohibited. Recordings made during the course would include statements, questions and comments made by students in the class, and these are not to be disseminated or repeated in any manner based on the recordings. Otherwise, please have cell phones turned off and put away while in lectures. *Thank you.*

Please have cell phones turned off and put away while in lectures.

Camosun College is a scent-free institution.

Please refrain from wearing scents. Thank you.